

# Aptamer-based biosensors

## nanoschematic

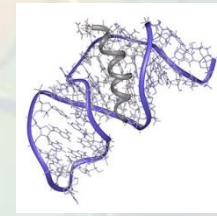
The concept of a nanoschematic is an extension of the schematic diagram, which is a graphical representation of a system. In the nanoschematic, the system is represented by a set of interconnected nodes and edges, where the nodes represent the components of the system and the edges represent the interactions between them. The nanoschematic is a powerful tool for analyzing and designing complex systems, as it allows for the visualization of the system's structure and the flow of information within it. The nanoschematic is also a useful tool for identifying potential problems and optimizing the system's performance.



## ***Aptamer-based biosensors***

- ***Qazvin university of medical science***
- ***Presented by: Seyed Majid Hosseini***
- ***Under supervision of Dr. Ahmadpour***

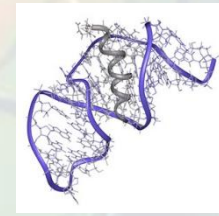
# CONTENTS



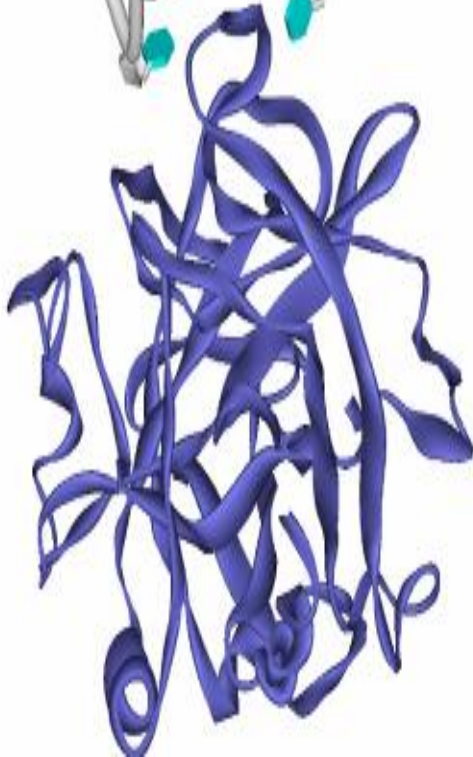
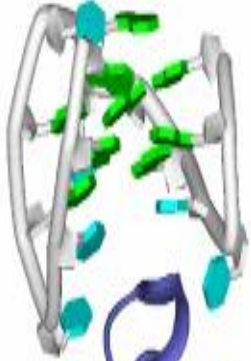
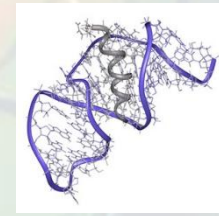
- **History and Background**
- **Introduction**
- **SELEX**
  - **Definition**
  - **SELEX cycle**
- **Biosensors**
- **Detection of Aptamer**
  - **Using affinity surfaces**
  - **Capillary electrophoresis**
  - **Flow cytometry**
- **Application**
  - **Aptamers in therapy**
  - **Aptamers in diagnostic**
- **Research Perspectives**
- **Conclusion**
- **References**



# History

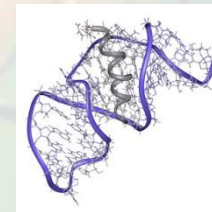


- **Spiegeman and his colleagues publish their work with nucleic acid in 1960.**
- **The result was the invention of PCR in 1986 by Molis.**
- **In 1990 SELEX (Selective expansion of ligands by exponential enrichment) were invented.**



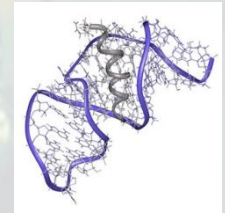
- ✓ **Aptamers** ( By Ellington) are oligonucleotides
- ✓ (DNA or RNA) that can bind with high affinity
- ✓ specificity to a wide range of target molecules such as drugs, proteins or other organic or inorganic molecules

Aptamers, derived from the Latin aptus, meaning “to fit”



- **Each** RNA molecules have **specific** sequences of nucleic acids.
- These molecules bind to different positions
- Establish hydrogen bonds and van der Waals molecules
- These RNA are called **Aptamer** and the binding site are named **Aptatop**.

# Aptamers VS Antibodies

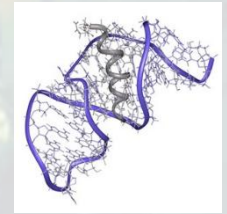


	<b>Aptamers</b>	<b>Antibodies</b>
<b>Affinity</b>	Low nM to pM	Low nM to pM
<b>Specificity</b>	High	High
<b>Production</b>	In Vitro Chemical Process	In Vivo Biological Systems
<b>Target Range*</b>	Wide: Ions, Cell, Toxins	Narrow: Immunogenics
<b>Batch to Batch Variation</b>	Little or No	Significant
<b>Chemical Modification*</b>	Easy	Limited
<b>Thermal Denaturation</b>	Reversible	Irreversible

**Aptamers:** Denser immobilization on the surface

**Aptamers:** Nuclease sensitive



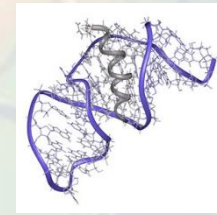


# Creation of Aptamers

- Aptamers are **generated** by an iterative selection process, called systematic evolution of ligands by exponential enrichment (**SELEX**).
- SELEX technology have extended aptamer selection from comparatively **simple mixtures of purified proteins** to whole **living cells**.
- SELEX **isolate** Aptamers that bind to specific target cells.

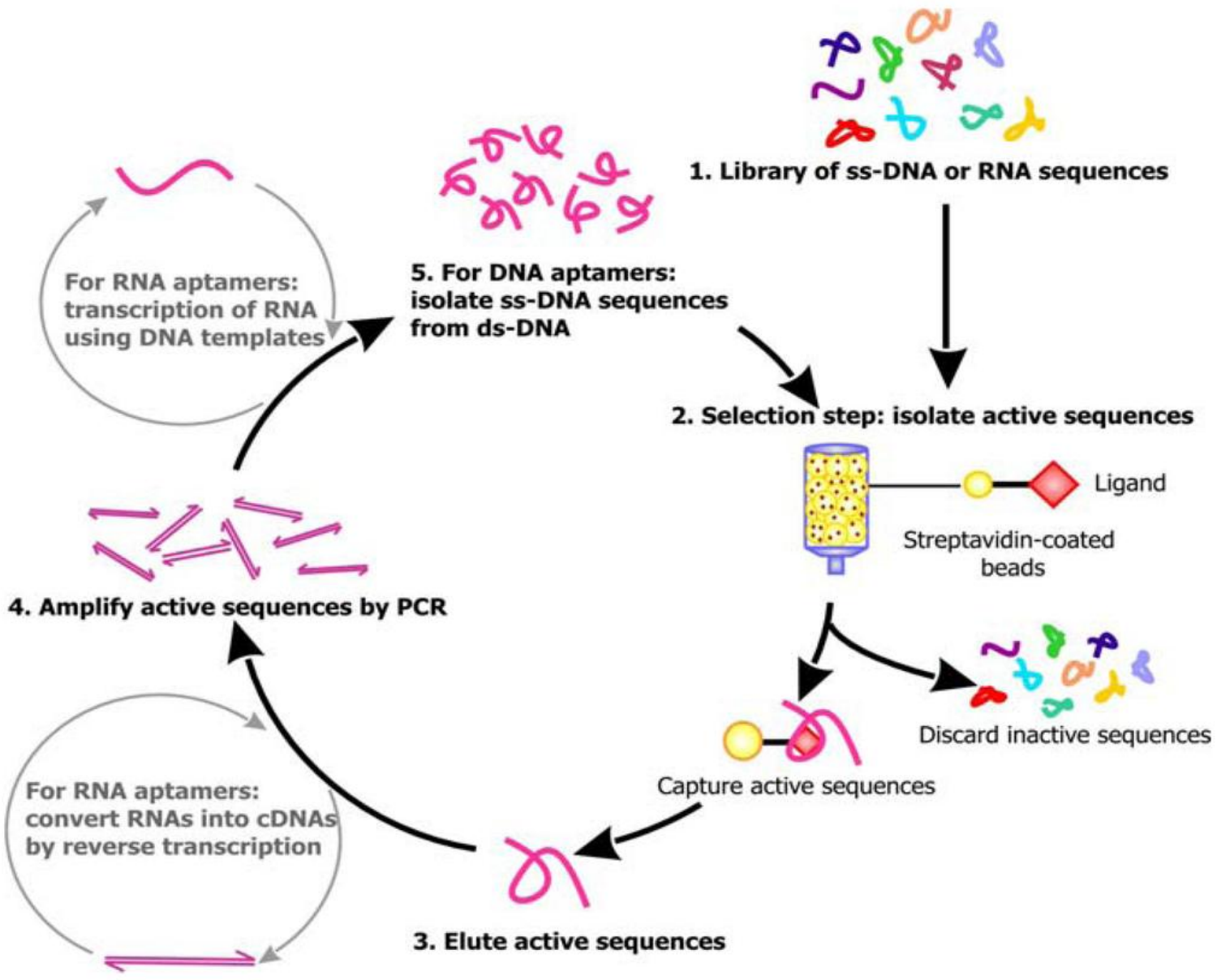


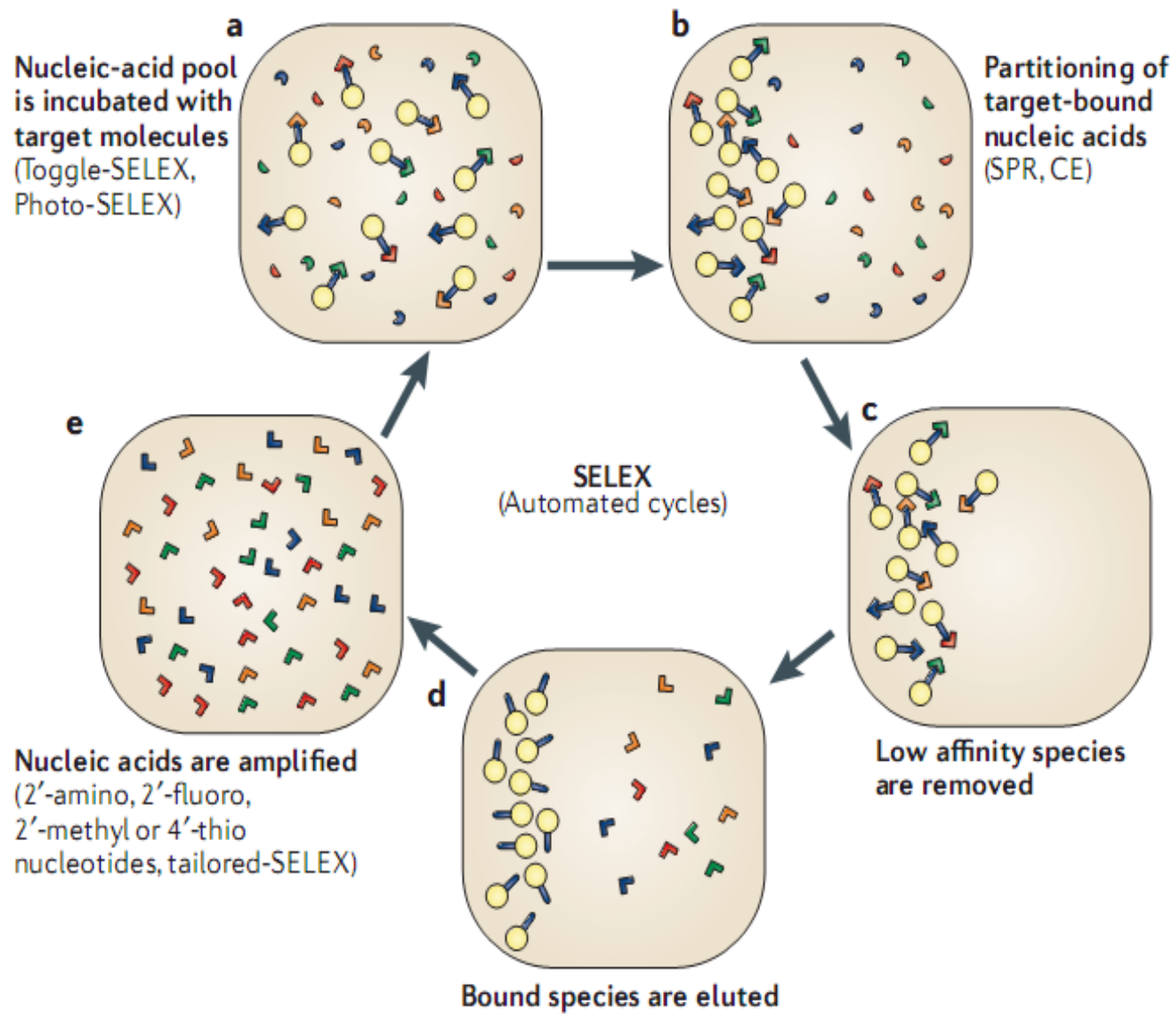
# SELEX



Systematic Evolution Of Ligands By EXponential Enrichment

- SELEX is a **multi-step** process in which strongly bind
- ligands are preferably selected by rounds of affinity assays and **PCR** amplification
- The SELEX method has permitted the identification of **unique RNA/DNA** molecules which bind to the target with very **high affinity** and **specificity**



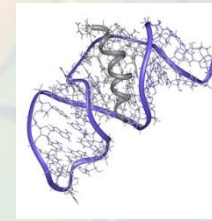




# circular dichroism spectroscopy



# Bio sensors



- A sensor is a device that transforms environmental information, ranging from the **concentration of a specific sample** component to total **composition analysis**, into an analytically useful signal.

- potentiometric
- amperometric
- potentiometric stripping analysis

- quartz crystal microbalance
- surface acoustic wave
- surface transverse wave

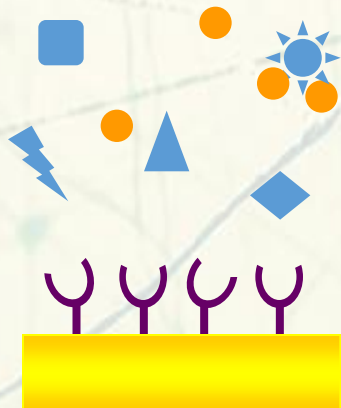
- absorbance
- luminescence
- fluorescence
- total reflectance fluorescence

## Recognition Part

- Antibody
- Enzyme
- Microorganism
- Aptamer

## Transducer

- Electrochemical
- Thermal
- Optical
- Mass Changes



Receptor

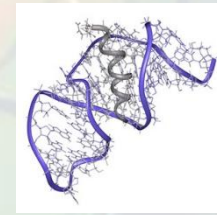


Transducer



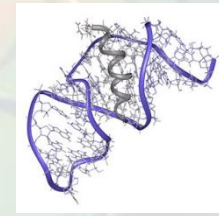
Recorder

# Essential Factors for Biosensors Operation



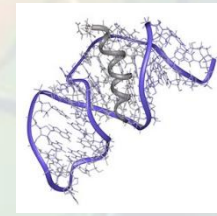
- *Sensitivity to presence of target*
  - The fabricated biosensor should show sensitivity to the presence of its own target.
- *Linearity of response*
  - The response of biosensor should be proportional to the concentration of its target.





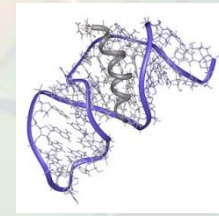
- *Selectivity*
  - Chemicals interference must be minimized for obtaining the correct result.
- *Reproducibility*
  - The response of the biosensor to the presence of target should be reproducible.

# Detection of Aptamer



- **Using affinity surfaces**
- **Using affinity tags**
- **Using column matrices or ligands**
- **Nitrocellulose membrane filtration**
- **Cross-linking**
- **Antibody-based**
- **Surface plasmon resonance**
- **Capillary electrophoresis**
- **Flow cytometry**
- **Automated selection**

# Using affinity surfaces



- **Affinity surfaces: allow proteins and small molecules to bind with them & have affinities with RNA or DNA.**
- **Magnetic beads, affinity titer plates**
- **RNA Aptamer against Panama influenza virus subtype A**



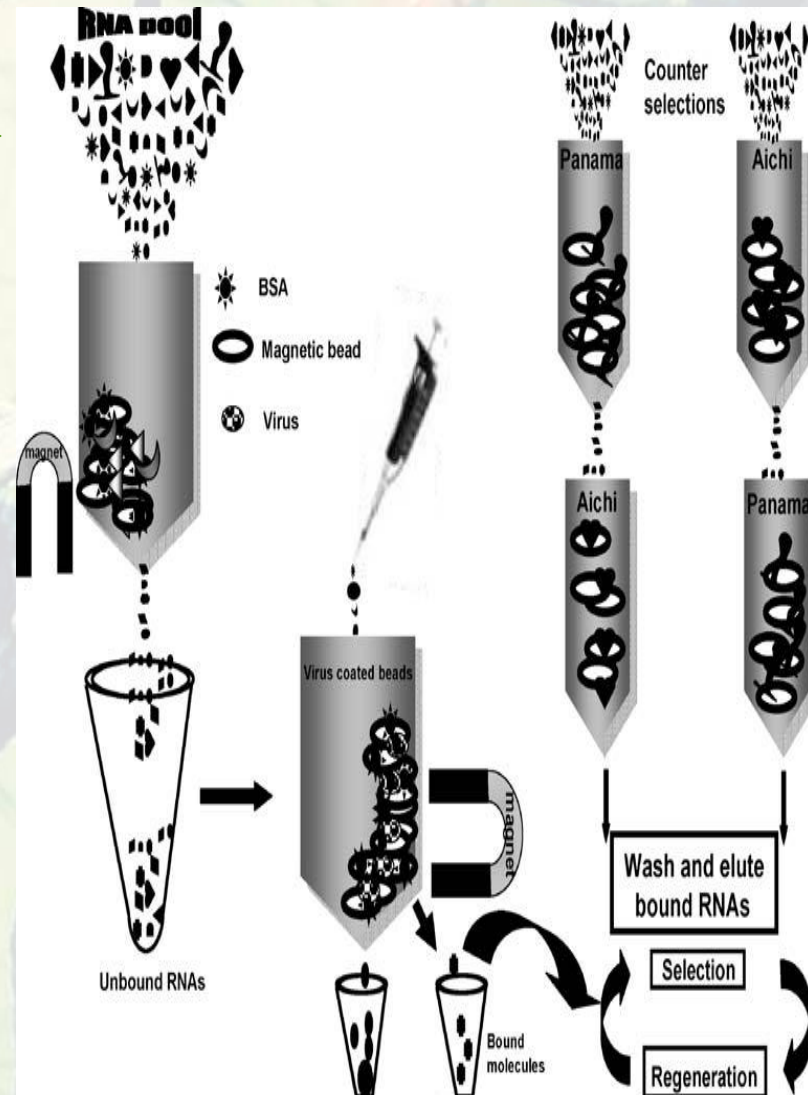
## Step 1

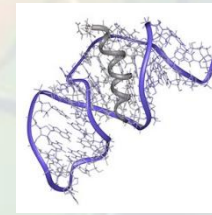
### Preparation

#### Coating and Amplification

- Coating the whole virus onto beads
- BSA Blocking
- Washing the coated beads.
- Denaturing the pool RNA(90 ° C for 2 min and allowed to cool at room temperature for 10 min)

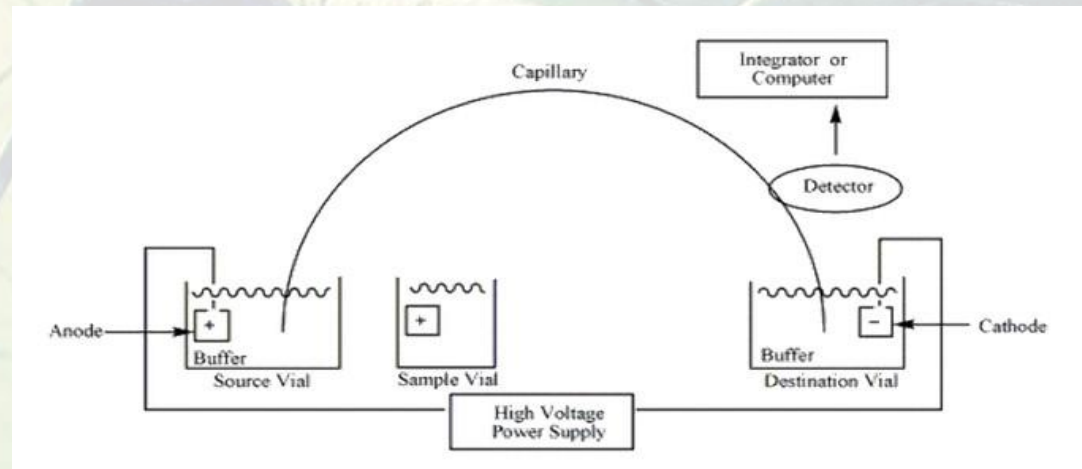
Bound molecules were precipitated by ethanol.

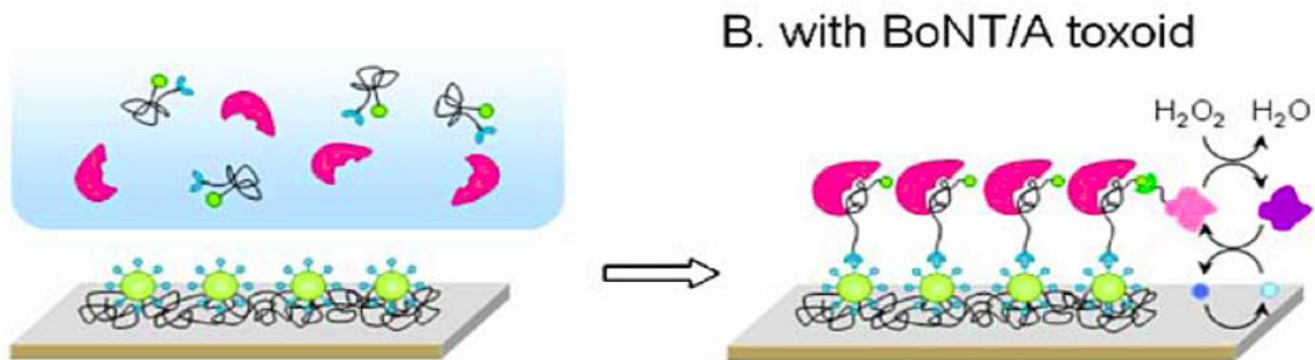
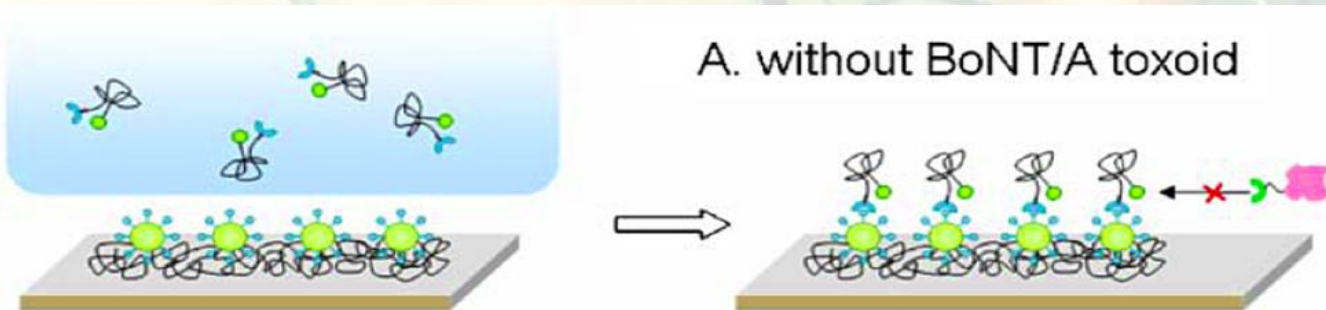




# Using capillary electrophoresis

- The nucleic acid sequences that bind the target undergo a mobility shift.
- no need to wash the active sequences off a column as in conventional SELEX, eliminating any kinetic bias.
- Higher speed, better resolution capacity, minimal sample dilution, fewer cycles.

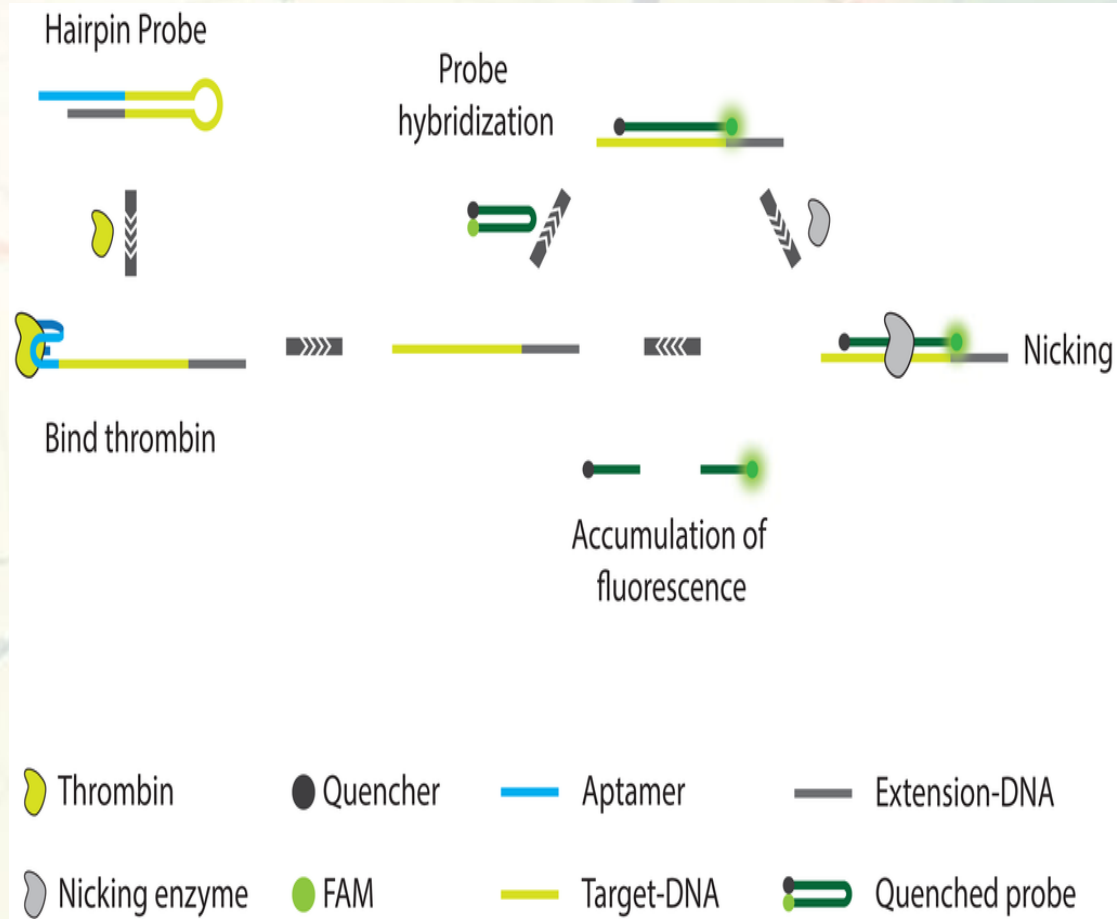
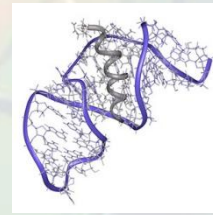




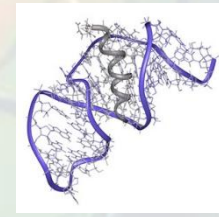
- |   |                        |  |                               |
|---|------------------------|--|-------------------------------|
|    | BoNT/A aptamer         |     | BoNT/A toxoid                 |
|  | Aptamer/toxoid complex |   | Streptavidin - dendrimer      |
|  | Oxidized TMB           |  | Reduced anti-fluorescein HRP  |
|  | Reduced TMB            |  | Oxidized anti-fluorescein HRP |



# Fluorescent signals



# Application



- Therapeutic Target
- Analytic Detection

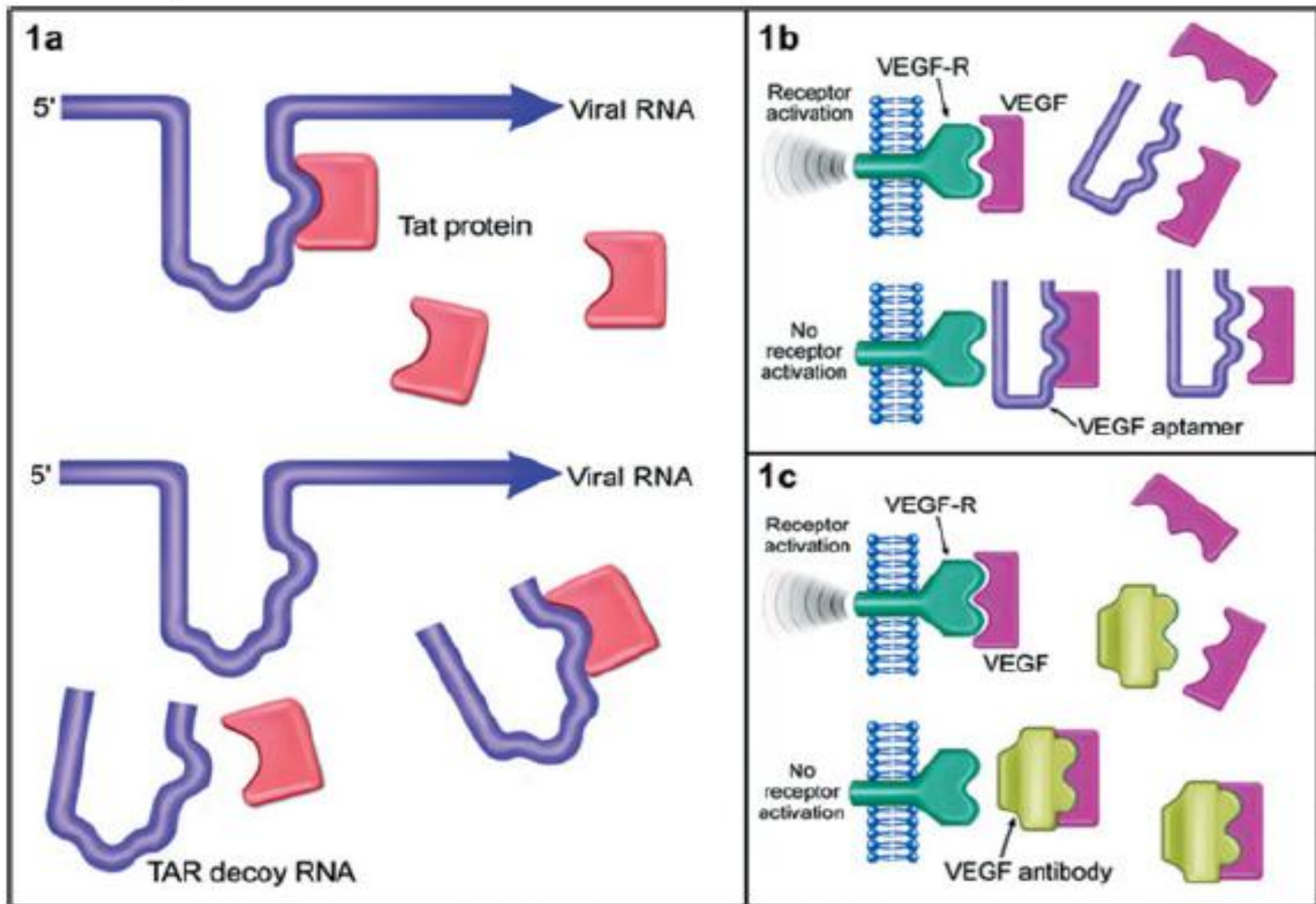
# APTAMER TO THERAPEUTIC TARGET

Target (alternative name)	$K_d$ (nM)	Therapeutic applications
$\alpha$ -thrombin	25	Prevent thrombosis
HIV-1 reverse transcriptase	1	Inhibit viral replication
HIV-1 Rev	<1	Inhibit viral replication
Fibroblast growth factor 2, basic	0.35	Prevent angiogenesis
Respiratory syncytial virus	40	Prevent infection
HIV-1 integrase	10	Inhibit viral replication
Vascular endothelial growth factor	0.14	Prevent neovascularization
Platelet-derived growth factor	0.1	Prevent tumour development
Immunoglobulin E	10	Prevent allergies
L-Selectin	3	Modulate inflammation
D-Adenosine	1,100	Unknown
Acetylcholine-specific auto-antibodies	60	Treat myasthenia gravis
Interferon- $\gamma$	68	Modulate inflammation and immune response
Keratinocyte growth factor	0.0002	Treat epithelial hyperproliferative disease
Neutrophil elastase	n/o	Modulate inflammation
P-selectin	0.04	Inhibit viral adhesion
Acetylcholine receptor	2	Control neurotransmission
Phospholipase $A_2$	118	Treat ARDS, septic shock
Protein tyrosine phosphatase	18	Inhibit oncogenesis, viral regulation
Activated protein C	110	Prevent thrombosis
CD4	0.5	Modulate immune response
Nuclear factor- $\kappa$ B	1	Treat chronic inflammatory disease
Lymphocyte function-associated antigen 1	500	Prevent tumour development, modulate inflammation
Cytoshesin 1	5	Modulate cytoskeletal reorganization
$\alpha$ v $\beta$ 3 integrin	2	Prevent tumour development

# APTAMER TO THERAPEUTIC TARGET

Target (alternative name)	$K_d$ (nM)	Therapeutic applications
Tenascin C	4	Prevent tumour development
Prostate-specific membrane antigen	2.1	Treat progressive malignant prostate disease
U1A	4.5	Modulate gene regulation
Gonadotropin-releasing hormone 1	50	Prevent tumour development
E2F transcription factor	15	Prevent tumour development
Neurotensin 1	1.5	Prevent viral infection
Factor IXa	0.65	Prevent thrombosis
NS3 protease	10	Treat hepatitis C virus infection
<i>Staphylococcus</i> enterotoxin B	420	Treat <i>Staphylococcus</i> infection
Chemokine (C-C motif) ligand 2 (MCP1)	3	Treat lupus
Angiopoietin 2	0.06	Prevent angiogenesis
HIV gp120	5	Inhibit viral infectivity
Calcitonin gene-related peptide	3	Treat migraine
HER3 (ERBB3)	45	Prevent tumour development
Cytotoxic T-lymphocyte-associated protein 4	10	Prevent tumour development
Cytohesin 2	115	Prevent tumour development
Nociceptin	110	Manage pain
Ghrelin	35	Prevent tumour development
Chemokine (C-X-C motif) ligand 10 (IP-10)	1.5	Modulate inflammation
Receptor tyrosine kinase RETC634Y	35	Prevent tumour development
Substance P	40	Prevent tumour development
Mucin 1	0.135	Prevent tumour development
Amylin	3	Treat pancreatic cancer
Angiopoietin 1	2.8	Prevent angiogenesis
Bovine prion protein	6.8	Treat prion and Alzheimer's disease
Plasminogen activator inhibitor 1	n/o	Prevent metastasis
Epidermal growth factor receptor variant III	33	Prevent tumour development



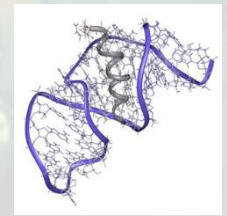


**Fig. 6.1** Aptamer–protein interaction. (a) TAR decoy RNA-mediated inhibition of Tat. The TAR decoy RNA is similar to the endogenous viral TAR RNA and assumes an analogous tertiary structure and in so doing, competes with viral TAR for Tat binding. (b) Anti-VEGF aptamer

inhibits VEGF receptor activation by VEGF by the same mechanism as (c) an antibody targeted to the same molecule. TAR, *trans*-activation response element; Tat, *trans*-activating regulatory protein; VEGF, Vascular endothelial growth factor.

# Aptamer and Epidermal growth factor receptor

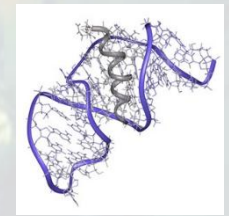
- Epidermal growth factor receptor (EGFR/HER1/c-ErbB1), is overexpressed in many solid cancers.
- TuTu22 aptamer was able to recognize cancer cells expressing EGFR but did not bind to the EGFR-negative cells.
- This DNA aptamers will facilitate the development of novel targeted tumor cell detection, cancer cell imaging and cancer therapy.



## analytical applications

- Affinity Chromatography
- Capillary Electrophoresis
- *In vitro* and *in vivo* diagnostic tools
- Targeting intracellular target molecules
- Drug Discovery
- Therapy
- Protein Purification
- Diagnostics
- ELISA
- Western Blotting



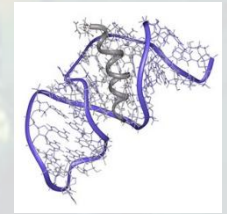


## Analytic Detection: Aptamers to Nucleic Acid

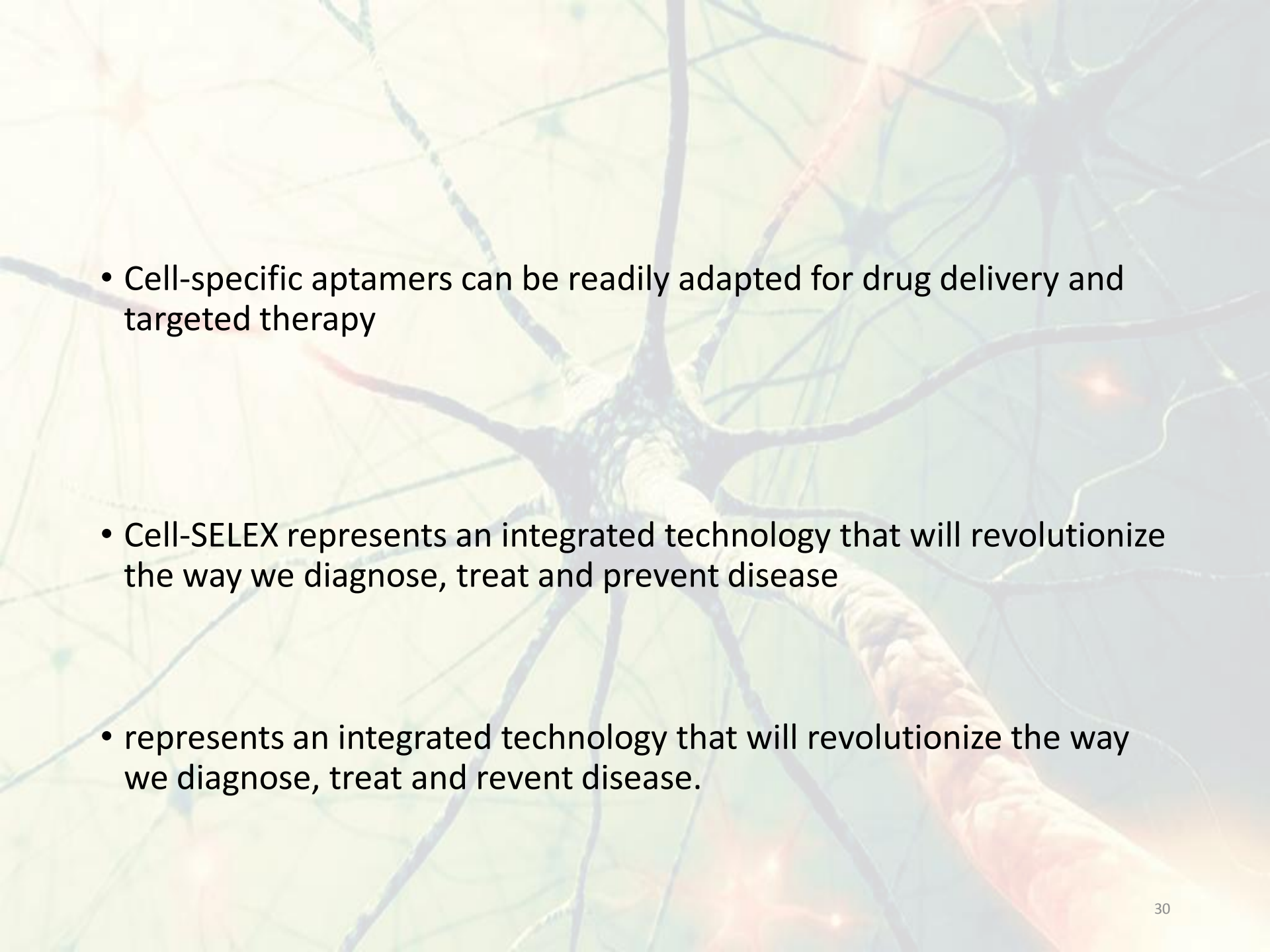
- ✓ The first aptamer to a nucleotide was isolated in 1993 by Sassanfar and Szostak
- ✓ In 2004 RNA selection aptamers that recognized the triphosphate of ATP were selected in the laboratory of Jack W. Szostak (Sazani et al., 2004)
- ✓ Interact with the part of the target molecule that faced the matrix, but did not strongly interact with the part that had greatest accessibility.

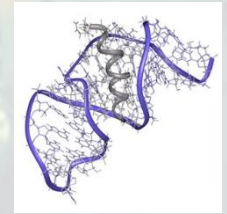


# Conclusion



- Aptamer technology has made significant strides.
- Aptamers can be applied to many fields of molecular medicine
- The identification of aptamer molecular targets constitutes a novel method of biomarker discovery

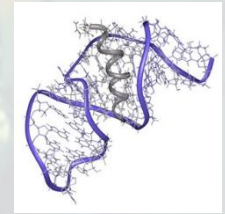
- 
- Cell-specific aptamers can be readily adapted for drug delivery and targeted therapy
  - Cell-SELEX represents an integrated technology that will revolutionize the way we diagnose, treat and prevent disease
  - represents an integrated technology that will revolutionize the way we diagnose, treat and prevent disease.



# Research Perspectives

- ✓ Aptamers are the very astonishing way to diagnose and diagnosis disease
- ✓ It seems using Aptamer will help us one step closer to understanding all sort of sickness especially in cancer world
- ✓ Limitation and weakness point of this knowledge is still under observation
- ✓ Also we still need more aptamer in order to diagnose more and specific illness.

# References



1. Vinkenburg JL, Mayer G, Famulok M. Aptamer-based affinity labeling of proteins. *Angewandte Chemie*. 2012;51(36):9176-80.
2. Zhu H, Li J, Zhang XB, Ye M, Tan W. Nucleic acid aptamer-mediated drug delivery for targeted cancer therapy. *ChemMedChem*. 2015;10(1):39-45.
3. Xiang D, Shigdar S, Qiao G, Wang T, Kouzani AZ, Zhou SF, et al. Nucleic acid aptamer-guided cancer therapeutics and diagnostics: the next generation of cancer medicine. *Theranostics*. 2015;5(1):23-42.
4. Zhou W, Huang PJ, Ding J, Liu J. Aptamer-based biosensors for biomedical diagnostics. *The Analyst*. 2014;139(11):2627-40.
5. Charoenphol P, Bermudez H. Aptamer-targeted DNA nanostructures for therapeutic delivery. *Molecular pharmaceuticals*. 2014;11(5):1721-5.
6. Zhang J, Liu B, Liu H, Zhang X, Tan W. Aptamer-conjugated gold nanoparticles for bioanalysis. *Nanomedicine (Lond)*. 2013;8(6):983-93.
7. Kim YS, Gu MB. Advances in aptamer screening and small molecule aptasensors. *Advances in biochemical engineering/biotechnology*. 2014;140:29-67



8. Liu, H.; Rajasekaran, A.K.; Moy, P.; Xia, Y.; Kim, S.; Navarro, V.; Rahmati, R.; Bander, N.H. Constitutive and antibody-induced internalization of prostate-specific membrane antigen. *Cancer Res.* 1998, 58, 4055–4060.
9. . Dhar, S.; Gu, F.X.; Langer, R.; Farokhzad, O.C.; Lippard, S.J. Targeted delivery of cisplatin to prostate cancer cells by aptamer functionalized Pt(IV) prodrug-PLGA-PEG nanoparticles. *Proc. Natl. Acad. Sci. USA* 2008, 105, 17356–17361.
10. . Wang, A.Z.; Bagalkot, V.; Vasilliou, C.C.; Gu, F.; Alexis, F.; Zhang, L.; Shaikh, M.; Yuet, K.; Cima, M.J.; Langer, R.; et al. Superparamagnetic iron oxide nanoparticle-aptamer bioconjugates for combined prostate cancer imaging and therapy. *ChemMedChem* 2008, 3, 1311–1315.
11. Bagalkot, V.; Zhang, L.; Levy-Nissenbaum, E.; Jon, S.; Kantoff, P.W.; Langer, R.; Farokhzad, O.C. Quantum dot-aptamer conjugates for synchronous cancer imaging, therapy, and sensing of drug delivery based on bi-fluorescence resonance energy transfer. *Nano Lett.* 2007, 7, 3065–3070.
12. Chu, T.C.; Marks, J.W., III; Lavery, L.A.; Faulkner, S.; Rosenblum, M.G.; Ellington, A.D.; Levy, M. Aptamer: Toxin conjugates that specifically target prostate tumor cells. *Cancer Res.* 2006, 66, 5989–5992.



Thank you